Research Article



Phytochemical Screening and Toxicity Studies of Aqueous Leaf Extract of Traditional Medicinal Plant *Cistus ladaniferus (L)*. in Mice and Rats

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Abstract

The purpose of this study is to evaluate the acute and sub-chronic toxicity of the aqueous leaf extract of *Cistus ladaniferus* (*C. ladaniferus*) in rats and mice.

Acute toxicity was studied in mice distributed in several lots. The six batches of mice received intraperitoneal (ip) doses corresponding to 200, 400, 600, 800, 1600, 3200 mg / kg of body weight for 14 days. Control mice received 0.9% NaCl. For sub--chronic toxicity the aqueous extract was administered orally to adult Wistar rats at different doses (500-1500 mg / kg / day, respectively, for 90 days). The animals are sacrificed. Blood samples are taken to study the parameters (hematological and biochemical) and the organs (liver, spleen, kidneys) are taken for histopathological analysis.

The results obtained show that the aqueous extracts of *C. ladaniferus* have a lethal dose LD50 of 831.76 ± 0.12 mg / kg of body weight. In all the parameters taken into account, no visible change was observed in the treated mice compared with the control groups. Sub-chronic toxicity results showed no clinical symptoms nor biochemical or histological alteration in rats treated.

The result indicates that intraperitoneal and oral administration of the aqueous extract of *C. ladaniferus* did not produce a significant toxic effect in mice and rats.

Keywords: *Cistus Ladaniferus*; Acute Toxicity Test; Sub-Chronic Toxicity Test; Aqueous Leaf Extract; Phytochemical Screening; Traditional Medicinal Plant

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Abbreviations: (DL50), Dose médiane; (I.P), Intra-péritonéale; (EDTA), Ethylenediamine tetra-acetic acid; (Hb), hemoglobin; (HT), hematocrit; (RBC), red blood cells; (GB), blood cells; (PLAQ), platelets. (CRE), creatinine; (AST), aspartate aminotransferase; (ALAT), alanine aminotransferase; (NaCl), Le chlorure de sodium;(LDH), lactate dehydrogenase ;(ALP),alkaline phosphatase.

Introduction

Human societies have been in close contact with their environment since the beginning of their training and used the ingredients of the environment to obtain food and medicines [1]. Medicinal plants have historically proved their value as a source of molecules with the rapeutic potential, and today still represent an important reservoir for the identification of new drug molecules [2].

Although herbal medicine to occupy a prominent place in the therapeutic arsenal, he use of medicinal plants is not insignificant, indeed, many plants are toxic and their misuse can be dangerous [3-5]. It is clear that in therapy, everything depends on the dose that remains a masterpiece in the therapeutic arsenal of medicinal plants. One of the problems of using herbal medicines is that in many cases, no defined doses are prescribed, often resulting in overdose [6]. *C. ladaniferus* is a plant belonging to the family Cisitaceae, represented by seven genera comprises 16 species, particularly prevalent in the Mediterranean [7]. *C. ladaniferus* is widespread in Portugal, Spain, Italy, Algeria, and Morocco [8]. It is a plant widely used in Western herbal medicine.

Studies have shown that *C. ladaniferus* has antifungal [9], antiviral, anti-inflammatory, gastro protective, and antitumor effects [10]. Flavonoids and tannins are the main compounds of *C.* ladaniferus, they are widely appreciated for their potentially beneficial effects on health, as antioxidants [11,12]. The aqueous extract of *C. ladaniferus* has antidiarrheal and anti-propulsive activity [13]. It has also been considered as a curative and preventive treatment against hypertension [14].

In order to identify the toxicological aspect of *C. ladaniferus*, we proceeded to the determination of the lethal dose (LD50) allowing knowing the smallest dose which administered at one time causes death of 50% of the animals. This study also makes it possible to specify the clinical signs of intoxication and to define the limits of tolerance to the product. In addition to this irreversible toxicity, we have studied the risks of latent or non-apparent sub-chronic toxicity, which makes it possible to search for and characterize the functional and / or pathological changes caused by a long-term treatment, thus highlighting the possible effect of possible accumulation of the substance and detecting the target organs by histopathological examination.

Material and Methods

Plant Material

C. ladaniferus was harvested in the Khénifra region during the period May-June 2014. All samples were shaded before use. Botanical identification and authentication of the plant material was done by Ms Leila NASIRI, Professor of the Faculty of Sciences of Meknes, Morocco. The plant material was dried in the shade and protected from moisture at room temperature for a few days before being crushed and stored until use.

Preparation of plant extracts

30 g of the powder of the *C. ladaniferus* leaves were boiled in 300 ml of distilled water for 15 minutes. After cooling, the extract is concentrated on a rotary evaporator. The yield of the extraction is 13.3% relative to the dry plant.

Phytochemical screening

We proceeded to search for the main chemical groups present in our aqueous extract. These generally sought after chemical groups are: alkaloids, sterols, triterpenes, saponins and polyphenolic compounds (tannins, flavonoids, quinones). Phytochemical analysis was performed according to the methods described by [15-17].

Experimental Animals

Adult healthy Swiss mice weighting 20–30 g aged two months were used. These mice come from the breeding center of the Institut Pasteur, Casablanca, Morocco. For the sub-acute toxicity, albino Wistar rats weighing 170–260 g were used. These animals were provided by the Laboratory Animals of the Faculty of Sciences, University of Moulay Ismail, Morocco.All animals were fed with standard laboratory diets, given water ad libitum and maintained under laboratory conditions.

Acute toxicity tests

The animals were divided into seven experimental groups of 10 animals each (5 males and 5 females). The 1st group served as a negative control, while 2nd, 3rd, 4th, 5th and 6th was considered as tested groups received intraperitoneally *C. ladaniferus* extract at the doses of 200, 400, 600, 800, 1600, 3200 mg / kg, respectively.

Animals were observed individually for general behavioral (activity, gait, tremor ...), body weight changes and mortality during the first 30 min, 1, 2, 4, 6 hours and once daily for 14 days after administration of the extract.

Then the mice were sacrificed and their organs subjected to macroscopic observation [18]. The mortality is recorded 1, 2, 24 hours, then 2 and 3 days after taking the aqueous extract. The LD50 was calculated by the method of [19].

Sub-chronic toxicity study

In the sub-chronic study, Wistar rats in four groups of 8 animals (4 males and 4 females) for each dose level of *C. ladaniferus* were used in these tests. Group I was

considered as control and the other two groups which were considered as tested groups received the plant extract at a dose of 500 and 1500 mg/kg body weight respectively for 90 days. Toxicity was evaluated in terms of corporal and organ weights, gross behaviour, haematological, biochemical characteristics and mortality. The organs (liver, spleen, kidneys) are removed, weighed and stored in Bouin's fluid for histopathological analysis [18,20,21].

Blood analysis

The blood sample was collected into ethylene diamine tetraacetic acid (EDTA) tubes for hematological and biochemical parameters. Sera samples were prepared by 10 min centrifugation of blood at 3500 rpm.

Histopathological study

After sacrificing the rats, the liver and the kidneys were removed and fixed in formalin. The next step is dehydration. This is usually done with a series of alcohols; say 70% to 95% to 100%.Each organ was sectioned, embedded in paraffin wax and then stained with hematoxylin-eosin and examined under a light microscope.

Statistical Analysis

All results are expressed as mean \pm SD. The comparison of the groups with the control is carried out using the Primer of biostatistic software. P-values less than 0.05 were considered to be significant.

Results

Phytochemical Screening

The phytochemicals are Tannins, flavonoids and saponines presented in the aqueous leaf extract of *C. ladani-ferus* (Table 1).

Table 1: Result of qualita	tive phytochemica	l composition of C.	ladaniferus (L) extract
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Phytochemical	Extract
Flavonoïdes	++
Tannins	++
Stérols et triterpènes	-
Quinones	-

Saponines	+
Alcaloïdes	-

Bioavailability key; - = not detected, + = present in low concentration, ++ = present in moderately high concentration

Acute toxicity study

In this part of the work, we undertook the study of the acute in vivo toxicity linked to the extract of *Cistus ladaniferus*, in order to determine the LD50, and to evaluate the clinical signs observed.

Preliminary tests of the aqueous leaf extract of *Cistus ladaniferus*, administered (IP) to mice at concentrations 200, 400, 600, 800, 1600, 3200 mg/kg, demonstrate the absence of toxicity of the plant extract up to the dose 400 mg/kg: 0% mortality after 14 days.

Note that from the concentration 600mg/kg, we observed clinical signs (tremors, hard stomach, accelerated breathing, rigidity of the hind limbs, loss of appetite, etc. (Table 2)

	Control	200mg/kg	400mg/kg	600mg/kg	800mg/kg	3200mg/kg
Respiration	N	Ν	Ν	N	N N Change	
Food intake	N	Ν	Ν	Ν	Ν	Change
Immobility	N	Ν	Ν	Ν	Ν	Lethargy
Rigidity of the hind limbs	NP	NP	NP	NP	Р	Р
Body weight	N	Ν	Ν	Ν	Ν	Change
Change in skin	NC	NC	NC	NC	NC	N.O
Eyes	N	Ν	Ν	Ν	Ν	N.O

Table 2: Evaluation of the clinical signs of the aqueous leaf extract of C. ladaniferus intraperitoneally administered to mice

N= normal, N.P= not present, P= present, N.C= not change, NO= not observed

These signs disappeared after 48 hours. And at the end of the test, all the surviving mice return to their normal state.





After intraperitoneal administration of the different doses (200, 400, 600,800, 1600, 3200 mg/kg), the LD50 is respected in mice at a dose of 831.76 ± 0.12 mg/kg (intraperitoneal route) (Figure1). Deaths of male and female mice are observed 36 and 48 hours after administration of the product.

On all the parameters taken into account, no visible change was observed in the treated mice compared to the control groups. According to our results, *C. ladaniferus* could be classified as low toxicity.

Sub-chronic toxicity study

Relative organ weight

Daily oral administration of the aqueous leaf extract of *C. ladaniferus* for 90 days did not produce any obvious symptoms of toxicity or mortality up to the highest dose level of 500 and 1500 mg / kg / day (Table 3).

Table 3: Effect of oral administration of the aqueous leaf extract of C. ladaniferus on the weight of rats

Doses (mg/kg)	wek1	wek2	wek3	wek4	wek5	wek6	wek7	wek8	wek9	wek10	wek11	wek12
0	170.1±27.2	178,22±27.2	181.7±26.6	184.9±27.2	189.7±27.2	194.5±25.4	198.9±27.2	200.1±27.2	201.2±27.2	202.8±27.2	203.5±29.2	202.8±27.2
500	209.2± 30.2	218.1±20.2	230.9±28.9	238.7±25.1	246.1±30.2	250.2±26.1	252.6±15.3	256.2±26.6	259.1±29.2	259.7±26.1	260.3±28.8	260.12±28.5
1500	196.8±28.2	221.32±28.2	224.3±26.1	226.34±26.2	229.2±26.2	230.2±26.9	234.7±32.2	236.1±27.2	236.7±20.2	238.0±28.2	239.9±32.0	235.0±28.2

The body and organs weight of the animals given *C. ladaniferus* extract once a day during 90 days did not show any significant differences compared with the control group.

Relative weight of liver, kidney and spleen

The effect of the extract on the weights of vital body organs is shown in Table 4. The body weight variation

of the rats of both sexes in all test and satellite groups was statistically not significant compared to that of their controls. The results revealed that, the vital organs such as liver, kidney, heart and spleen were not adversely affected throughout the treatment by extract. Studies have shown that weight reduction of the body and internal organs is considered as an index of sensitivity to toxicity after exposure to a substance [22,23].

Table 4 : Organ weights (g) of the rats in the sub-chronic toxicity studies of the aqueous leaf extract of <i>C. ladaniferus</i> . n = 8, Values are ex-
pressed as mean ± SEM, * p >0,05

	Liver	Kidneys	Spleen
Control mg/kg	7.20 ± 0.7	0.61±0.02	$0.51.\pm0.04$
500mg/kg	6.87±0.30	0.62±0.02	0.53±0.05
1500mg/kg	6.67±0.19	0.59±0.03	0.50±0.03

The values are expressed as mean ± standard deviation of 8 rats in each group. Group I: Control (0 mg / kg); Group II: 500 mg / kg; Group III: 1500 mg / kg.

Hematological analysis

The effects of extract on hematological parameters are summarized in (Table 5). No significant difference was observed from the results of the hematological profile hemoglobin, hematocrit, red blood cells, white blood cells and platelets) of rats ofboth sexes. There were generally no significant differences noted between control and treated groups for the hematological parameters measured.

Parameters	Doses (mg/kg)			
	Control	500	1500	
Red blood cells (10 ⁶ /µl)	7,70±0,53	7,71 ±0.20*	7,93±0,40 *	
White blood cells ($10^3/\mu$ l)	9,10± 0,51	9,08± 0,15 *	8,95±0,59 *	
Haemoglobin (g/dl)	15,89± 0,85	15,43± 0,69 *	15,66 ±0,62 *	
Hématocrit (vol %)	40,87 ±0,45	41,75±0,66 *	41,62±0,8 *	
Platelets (10 ⁴ /µl)	70,97± 0,86	70,74±0,8 4*	70,99±0,74 *	

Table 5: Study of the haematological parameters of rats after daily oral administration of the aqueous leaf extract of *C. ladaniferus* for 90days. n = 8, Values are expressed as mean \pm SEM, * p >0,05

Values are expressed as mean ± standard deviation of 8 rats in each group.Group I: Control (0 mg / kg); Group II: 500 mg / kg; Group III: 1500 mg / kg.

Biochemical analysis

The effects of repeated administration of C. ladani-

ferus extract for 90 days on serum biochemical parameters are summarized in Table 6.

Table 6: Study of the biochemical parameters of rats after daily oral administration of the aqueous leafextractof C. ladaniferus . n = 8, Values
are expressed as mean ± SEM, * p >0,05

Parameters	Doses (mg/kg)				
	Control	500	1500		
Creatinine (µmol/l)	59±0.91	60,4±0.20 *	61,4±0.55 *		
ASAT (U/L)	83,5± 6,52	80,1±6,56 *	84,75±3,19 *		
ALAT (U/L)	22,1± 3,87	23,2± 1,28 *	23,5±1,30 *		
Total protein (g/l)	82,01±7,41	84,2±7,25 *	82,2±7,48 *		

Values are expressed as mean ± standard deviation of 8 rats in each group. Group I: Control (0 mg / kg); Group II: 500 mg / kg; Group III: 1500 mg / kg.

All the tested biochemical parameters including Creatinine (CRE), aspartate aminotransferase (AST), alanine aminotransferase (ALAT), and total proteins were within normal limits compared to control group.

No significant changes were observed in the different biochemical parameters studied. Normal creatinine values suggest that this extract did not alter renal structure and function. In our study and following the treatment of the rats, no adverse effects were observed on the morphology of the kidneys, spleen and liver of the treated rats.

Histology of liver, spleen and kidney

Histology of the kidney, spleen and liver sections of male and female rats who received *C. ladaniferus* extract at different doses for 90 consecutive days is presented in Figure2, Figure 3, and Figure 4. Microscopical analysis of the examined internal organs displayed normal aspect of the liver, kidney and spleen for the two doses used (500mg / kg and 1500 mg / kg). No significant differences were grossly detected between control and experimental groups.



Figure2: Histological section of male rat kidney (A) and female rat (B) control. Male rat (C) and female rat (D) treated (dose of 500mg / kg). Male rat (E) and female rat (F) treated (dose of 1500mg / kg). Vascular congestion. Hematoxylin Eosin Stain (Average magnification)





Figure 3: Histological section of male rat spleen (A) and female rat (B) controls. Male rat (C) and female rat (D) rats treated (500 mg / kg dose) treated male rat (E) and female rat (F) rats (dose of 1500 mg / kg). Eosin hematoxylin staining. (Average magnification)



Figure 4: Histological section of male rat liver (A) and female rat (B) controls. Male rat (C) and female rat (D) treated (dose of 500mg / kg). Male rat (E) and female rat (F) treated (dose of 1500mg / kg). Eosin hematoxylin staining. (Average magnification)

Discussion

Toxicological studies of plants with therapeutic uses are today a key step in the process of valuing aromatic and medicinal plants.

Conventionally, in the presence of a substance endowed with biological activity, the study of the toxicity and in particular the evaluation of the LD50 proves essential. This value already gives an estimate of the toxicity of the studied substance and will be used to establish the therapeutic coefficient characterized by the ratio between the LD50 and the active dose.

Phytochemical analysis of the aqueous extract of *C. ladaniferus* revealed the presence of flavonoids, tannins and saponins.

Acute toxicity assessment allowed determination of LD50 in mice treated intraperitoneally with aqueous extract of *C. ladaniferus*. The value of the LD50 was estimated by the Miller and Tainter graphical method. The results obtained show that the aqueous extract of leaves of *C. ladaniferus* has a lethal dose LD50 of 831.76 \pm 0.12 mg / kg.

According to the [24] classification, chemicals

with an LD50 between 500 and 5000 and between 5000 and 15000 mg / Kg are considered almost moderately and nontoxic, respectively. From this classification it can be concluded that the aqueous leaf extract of *C. ladaniferus* was classified as moderately toxic in mice. A study by [13] showed that the aqueous leaf extract of *C. ladaniferus* leaves is considered non-toxic. Another study by [9] showed that the aqueous leaf extract of *C. ladaniferus* is considered moderately toxic.

The evaluation of sub-chronic toxicity at doses of 500 and 1500 mg / kg / day by daily oral administration of the aqueous leaf extract of *C. ladaniferus* resulted in no death, no evidence of change in morphology, body weight and body weight. The behavior of treated animals is quite comparable to that of untreated rats. Studies have shown that reduction in body and internal organ weight is considered an index of sensitivity to toxicity after exposure to a toxic substance [22,23]. Our aqueous leaf extract of *C. ladaniferus* was also without effect on the biochemical and hematological parameters. Normal creatinine values suggest that this extract did not alter renal structure and function.

With respect to other biochemical parameters such as total protein no significant difference was observed compared to the control group. In addition, the transaminase enzymes SGOT (AST) and SGPT (ALT) were not disturbed, the liver was not reached. These parameters are not statistically different (p> 0.05).

In addition, that transaminases enzyme SGOT (AST) and SGPT (ALT) were not disturbed, the liver was not reached and the total proteins did not undergo variations during the study. These parameters are not statistically different (p> 0.05).

Transaminases, LDH, and alkaline phosphatases ALP are good indicators of liver, heart and kidney toxicity, respectively [25]. The microscopic observation of these organs shows a normal appearance of the kidneys of the rats and this for the two doses used (500mg / kg and 1500 mg / kg). The liver also has a normal appearance similar to that of control rats.

Histological sections of kidneys, spleen, and liver from treated rats appeared normal compared to control rats (Figure 2, Figure 3 and Figure 4). Hepatic tissue had liver lobules formed from normal hepatocyte cells in the male and female treated groups (Figure 4). Our results are in agreement with those found by [26,5].

The liver and kidneys are target organs for toxic chemicals because of their essential functions in body detoxification and excretion processes [26,27]. As well as the liver is considered an important organ in a toxicological study [28]. Histological features of nothing showed a normal structure of glomeruli and Bowman capsules (Figure 2). Thus, the morphology of the proximal and distal convoluted tubules shows no abnormality. Similar results have been reported by [5] in rats treated with Senna alata.

Histopathological examination of the spleen sections showed a normal splenic parenchyma with no abnormalities in the white and red pulp of both treated and control groups (Figure 3). Our results are in agreement with those found by [29]. The present study shows that daily oral administration for 90 days of the aqueous leaf extract of *C. ladaniferus* does not cause any mortality, nor alter the biochemical and histopathological parameters of the treated rats. Under these experimental conditions, the plant extract appears to be devoid of toxicity in male and female albino rats.

The toxicological study of medicinal plants is a substantial step in order to determine the limits of their tolerances and subsequently to optimize their use. The information from the set of results presented in this work on the experimental acute toxicity data in mice suggests that the aqueous extract of *C. ladaniferus* administered intraperitoneally is classified as a weakly toxic. However, sub-chronic toxicity shows that the aqueous extract of *C. ladaniferus* is completely devoid of toxic effects. In this sense, the rare work done on the toxicity of *C. ladaniferus* confirms the safety of the plant. This issue is currently of great interest and deserves to be widely exploited.

Conflict of interest statement

We wish to confirm that there are no known conflicts of interest associated with this publication and there has been no significant financial support for this work that could have influenced its outcome.

References

1. Jamshidi-Kia F, Lorigooini Z, Amini-Khoei H (2018) Medicinal plants: Past history and future perspective. Journal of Herbmed Pharmacology 7: 1-7.

2. Atanasov et al. (2015) Discovery and resupply of pharmacologically active plant-derived natural products: A review. Europe PMC Funders Group Author Manuscript Biotechnol Adv.Author manuscript; available in PMC 33: 1582–614.

3. Balajis S, Chempakam B (2010) Toxicity prediction of compounds from turmeric (Curcuma longa L). Food Chem Toxicol 48: 2951-9.

4. Santosh N, Mohan K, Royana S, Yamini TB (2010) Hepatotoxicity of tubes of indian Kudzu (Pueraria tuberrosa) in rats. Food Chem Toxicol 48:1066-71.

5. Roy S, Ukil B, Lyndem LM (2016) Acute and sub-acute toxicity studies on the effect of Senna alata in Swiss Albino mice. Cogent Biology 2: 1272166.

6. Usman MM, Sule MS, Gwarzo MY (2014) Toxicological studies of aqueous root extract of Euphorbia lateriflora (Schum and Thonn) in rats. Journal of Medicinal Plants Studies 2.

7. Talavera S, Gibbs PE,Et Herrera J (1993) Reproductive Biology Of Cistus Ladaniferus (Cistaceae). Plant Syst. And Evol 186: 123-34.

8. Mariotti JP, Tomi F, Casanova J, Costa J, Bernardini AF (1997) Composition of the Essential oil of Cistus ladaniferus L. Cultivated in Corsia (France), Flavour and Fragrance Journal 12: 147-51.

9. Barros L, et al. (2013) Antifungal activity and detailed chemical characterization of Cistus ladanifer phenolic extracts. Industrial Crops and Products 41: 41–5.

10. Skoric M et al. (2012) Cytotoxic activity of ethanol extractsof ethabol of in vitro grownCistus creticus subsp. Creticus L. on human cancer cell lines, industrical Crops and products 38 : 153-9.

11. Amensour M, Sendra E, Pérez-Alvarez JA, Skali-Sen-

haji N, Abrini J, Fernández-López J (2010) Antioxidant activity and chemical content of methanol and ethanol extracts from leaves of rockrose (C. ladaniferus). Plants Foods Hum. Nutr 65: 170-8.

12. Guvenc A, Yildiz S, Ozkan AM, Erdurak CS, Coskun M, Yilmaz G, Okuyama T, Okada Y (2005) Antimicrobiological studies on Turkish Cistus species. Pharm. Biol 43: 178-83.

13. Aziz M. et al. (2011) Relaxant effect of aqueous extract of Cistus ladaniferus on rodentintestinal contraction, Fitoterapia 77: 425-8.

14. Belmoukhtar M, et al. (2009) Antihypertensive and endothelium-dependent vasodilator effects of aqueous extract of Cistus ladaniferus. Biochemical and Biophysical Research Communications 389: 145-9.

 Wagner H, Bladt S (1996) Plant Drug Analysis: A Thin Layer Chromatography Atlas. Berlin. Springer Science Business Media.

16. Bekro YA, Bekro JAM, Boua BB, Tra Bi FH, Ehilé EE (2007) Etude ethnobotanique et screening phytochimique de Caesalpinia benthamiana (Baill.) Herend et Zarucchi (Caesalpiniaceae). Rev. Sci.Nat 4: 217-25.

17. Trease E, Evans WC (1987) Phermacognosy. Billiare Tindall. London 13: 61-2.

18. Kanjanapothi et al. (2004) Toxicity of curde rhizome extract of Kaempferia galanga L (Proh Hom). Journal of Ethno pharmacology.

 Miller LC, Tainter ML (1944) Estimation ofLD50 and its error by means of log-probit graph paper. Proc Soc Exp Bio Med 57: 261.

20. Palmeiro NMS, Almeida CE, Ghedini PC, Goulart LS, Pereira MCF, Huber S, Silva JEP, Lopes S (2003) Oral subchronic toxicity of aqueous crude extract of Plantago australis leaves. Journal of ethno pharmacology 88: 15-8.

21. Al Habori M, Al-Aghbari A, Al-Mamary M, Baker M (2002) Toxicological evaluation of Catha edulis leaves: a long term feeding experiment in animals. Journal of Ethno pharmacology 83: 209-17.

22. Raza M, Al-Shabanath OA, El-Hadiyah TM, Al-Majed AA (2002) Effect of prolonged vigabatrin treatment on hematological and biochemical parameters in plasma, liver and kidney of Swiss albino mice. Scientia Pharmaceutica 70: 135-45.

23. Thanabhorn S, Jenjoy K, Thamaree S, Ingkaninan K., Panthong A (2006) Acute and subacute toxicity study of the ethanol extract from Lonicera japonica Thunb. Journal of Ethno pharmacology 107: 370-3.

24. Loomis TA, Hayes AW (1981) Loomis's essentials of toxicology. New York: Academic Press 1996.

25. Martin DW, Mayes PA, Rodwell YM. In:Harper's Review of Biochemistry. 18th edn, Lange Medical 61.

26. Bello I, Bakkouri AS, Tabana YM, Al-Hindi B, Al--

Mansoub, MA, Mahmud R, Asmawi MZ (2016) Acute and Sub-Acute Toxicity Evaluation of the Methanolic Extract of Alstonia scholaris Stem Bark. Med. Sci 4: 4.

27. Adekomi DA, Musa AA, Tijani AA, Adeniyi TD, Usman B (2011) Exposure to smoke extract of Datura stramonium leaf: Some of its effects on the heart, liver, lungs, kidneys and testes of male Sprague Dawley rats. Journal of Pharmacognosy Phytother 3: 67-75.

28. Kalaiselvia P, Umaa M, Suresha M, Thulasiramana K, Lakshmidevib E (2013) Chronic toxicity studies of aqueous leaf extract ofIndian traditional medicinal plant Ocimum tenuiflorum (Linn.) in rats. European Journal of Experimental Biology 3: 240-7.

29. Treadway S (1998) An ayurvedic herbal approach to a healthy liver. Clinical Nutrition Insights 6: 1-3.

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